

<https://doi.org/10.59674/pbk2>

PHYTOCHEMISTRY, MEDICINAL AND NUTRITIONAL IMPORTANCE OF *ASPARAGUS RACEMOSUS*

Kashif JAVAID¹, Shabbir HUSSAIN^{2*}, Shahzada Khurram SYED³,
Iqbal AHMAD⁴, Shazma MASSEY⁵, Amina ASGHAR⁶, Israr AHMAD²,
Muhammad AMJAD¹, Aqsa RUKHSAR¹, Maleeha HAFEEZ¹

¹Department of Chemistry, Lahore Garrison University, Pakistan

²Institute of Chemistry, Khwaja Fareed University of Engineering and Information
Technology, Rahim Yar Khan, Pakistan

³Department of Basic Medical Sciences, School of Health Sciences,
University of Management and Technology, Lahore, Pakistan

⁴Department of Chemistry, Allama Iqbal Open University, Islamabad Pakistan

⁵Department of Chemistry, Forman Christian College (A Chartered University),
Lahore, Pakistan

⁶Department of Chemistry, Division of Science and Technology,
University of Education, Lahore, Pakistan

Summary: Plants possess very important phytochemical constituents and are rich in nutritional and medicinal contents. Current studies were performed to review the phytochemistry, nutritional and pharmaceutical value of *Asparagus racemosus* (shatavari). This plant contains numerous steroidal saponins, steroids, alkaloids, flavonoids, cyclic hydrocarbons, tannins, anthraquinones, polysaccharides, oligospirostanoside, trace minerals, derivatives of dihydrophenanthrene and derivatives of furans and essential oils. *A. racemosus* has shown neuroprotective, antitussive, immunomodulatory, antidepressant, nootropic, anti-amnesic, anti-ulcer, antiparasitic, antidiabetic, anticancer, aphrodisiac, antibacterial, anti-inflammatory, anti-diarrheal, antioxidant, hepatoprotective, hypocholesterimic, antirolithiatic, anti-sebum and wound healing effects. It has been found effective in improving sperm count and also used to treat blood dysentery, skin problems, kidney stones, dysentery, diarrhoea, epilepsy, general weakness, leucorrhoea, gynecological disorders, lactation issues and teratological disorders. Its root extract is an important ingredient of many useful drug formulations including Abana, Diabecon, EyeCare, Geriforte, Himplasia, Lukol and Renalka. Its roots are rich in many nutrients and can be orally given to dairy animals as food supplements to increase the milk production, reproduction capacity and immune system. Shatavari extracts can be used to increase the nutritional value of bakery products and fortification of milk.

Keywords: *A. racemosus*, phytochemistry, medicinal, drugs, nutritional

INTRODUCTION

The plants are largely investigated by researchers due to their tremendous nutritional [1-3] and medicinal [4-6] value associated with phytoconstituents [7-9]. *Asparagus racemosus* is a famous medicinal plant which is employed to treat a wide range of medicinal issues including and urinary tract infections, leucorrhoea, benign prostatic hyperplasia, anxiety disorders, dysmenorrhea, angina, hypertension and hyperlipidemia [10]. This plant is generally called as "shatavari" (shat: "hundred"; variety: "curer") since it was used for the treatment of numerous diseases especially the issues related to female reproduction and was known as "a Female Tonic". The plant belongs to genus *Asparagus* which have about 300 species. *A. racemosus* is the most cultivated species for medicinal uses [11, 12]. It belongs to the species *Asparagus racemes*, genus *asparagus*, subfamily *asparagoideae*, family *asparagaceae*, order *asparagales* and class *angiosperms* [13]. The plant is 1-2 meter height having beautiful and widespread branches and extensive tuberous root system [14]. *A. racemosus* is found at lower altitude and in tropical and subtropical regions of Australia, Africa and Asia [14, 15]. Each part of this plant are rich in medicinal contents but roots are top of the list [14].

This review elaborates the phytochemistry, nutritional and pharmacological effects of *Asparagus racemosus* in detail.



FIGURE 1 *Asparagus racemosus* (shatavari) [16]

PHYTOCHEMICAL SCREENING

A. racemosus contains a variety of secondary metabolites, including alkaloids, derivatives of dihydrophenanthrene, furan compounds, steroids, flavonoids and essential oils. However, the chief components of *A. racemosus* are steroidal saponins, which owe numerous biological activities to the plant [10]. Some important phytochemicals are described below:

- The plant roots have been found to contain the following steroidal components: 3-O-[α -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranosyl-(1 \rightarrow 4)-O- β -D-glucopyranosyl]-25(S)-spirosta-3 β -ol [17], Shatavarins [18], Asparanin A [19], Immunoside [19], (1S,2R,3S,8S,9S,10S,13S,14S,16S,17R,22R,25R)-21-nor-18 β , 27 α -dimethyl-1 β , 2 β ,3 β -trihydroxy-25-spirost-4-en-19 β -oic acid [20], Sarsasapogenin [21], Diosgenin [21], Sitosterol [22-24], Anti-HIV compounds [25], Filiasparoside C [19, 26], Shatavaroside A [26] and Shatavaroside B [26].
- The roots also contain two alkaloids namely Asparagamine A [27], Polycyclic alkaloid [28].
- Its roots were found to contain flavonoids including 8-Methoxy-5,6,4-trihydroxyisoflavone-7-O- β -D-glucopyranoside [29]. The tuberous roots (woody parts) also possessed Cyanidine-3-galatoside [21] and Kaempferol [21].
- The leaves of *A. racemosus* contain flavonoids namely 5-hydroxy-3,6,4'-trimethoxy-7-O- β -D-glucopyranosyl-[1 \rightarrow 4] -O- α -D-xylopyranoside [30] and Quercetin-3-glucuronide [30].
- The flowers and fruits of the plant contain flavonoids including Quercetin [30], Rutin [30] and Hyperosid [30].
- Steroidal component namely Racemoside A, B, C was isolated from defatted fruits [31].
- The roots contain Racemosol (9, 10-dihydro-1, 5-dimethoxy-8-methyl-2, 7-phenanthrene diol) [32] and Racemofuran [33] which are derivatives of Dihydrophenanthrene and Furan.

A. racemosus contains steroidal saponins, also called shatvarins [34, 35], polycyclic alkaloid-asparagamine A (a cage type pyrrolizidine alkaloid) [36], cyclic hydrocarbon-racemosol (dihydrophenanthrene) [32], carbohydrates-polysaccharides (mucilage) [37], isoflavones-8-methoxy-5, 6, 4-trihydroxy isoflavone-7-O-beta-D-glucopyranoside [29], racemofuran [33], oligospirostanoside (also called immunoside) [38], essential fatty acids (such as quercetin 3-glucourbnides, diosgenin, vitamin A and gamma linoleinic acids) [39], undecanyl cetanoate and sitosterol (present in roots) [23]. The woody portion of tuberous roots contains sarsapogenin and kaepfrol [40]. Fruits and flowers are rich in flavonoids which are glycosides of hyperoside, rutin and quercitin [41]. The roots contain trace mi-

nerals including Zn (53.15), Co (22.00 mg/g), Mn (19.98 mg/g), Cu (5.29 mg/g), along with selenium, potassium, magnesium and calcium [42].

An isoflavone namely 8-methoxy-5,6,4'-trihydroxyisoflavone-7-O- β -d-glucopyranoside was obtained from root extracts of *A. racemosus* [43]. Ethanolic extract of *A. racemosus* roots was found to contain a derivative of 9,10-dihydrophenanthrene namely racemosol (9,10-dihydro-1,5-dimethoxy-8-methyl-2,7-phenanthrenediol) [44], kaempferol, a polycyclic alkaloid known as asparagamine and steroidal saponins such as shatavarins (I, IV) while the presence of sarsasapogenin was shown in its roots, leaves, and fruit extracts. The asparanin (A,B,C) and adscendin (A, B) were also explored from the ethanolic extracts of *A. racemosus* roots [45]. The methanolic extracts of *A. racemosus* fruits were analyzed (by spectroscopy and various chemical procedures) for the following 3 steroidal saponins [46]: Racemoside A, Racemoside B and Racemoside C.

Isolation and structural elucidations were done by reversed phase high-performance liquid chromatography and NMR (1D and 2D), respectively for novel steroidal saponins and shatavarin V from the *A. racemosus* roots [47]. Various spectroscopic techniques were used for isolation and structural elucidation of a valuable sarsasapogenin glycoside from *A. racemosus*; this glycoside is known as immunoside ($C_{45}H_{74}O_{16}$) and has a significant potential of altering immune system [48]. Root extracts showed a number of total phenolic contents, total flavonoids [49], glycosides, saponins, anthraquinones, tannins, carbohydrates, phytosterols and steroids [50]. By NMR and QTOF-MS studies, filiasparoside C and two new steroidal saponins (shatavarosides A and B) were separated and structural elucidations were made [51]. An important antioxidant compound "racemofuran" was isolated from *A. racemosus* roots by TLC [52]. Different parts of *A. racemosus* have also shown valuable amounts of Cu, Zn, Mn, Cu, Fe, K, Mg and Ca [53].

PHARMACOLOGICAL VALUE

Neuroprotective Effects: Alzheimer's disease and Parkinson's diseases are caused by oxidative stress and excitotoxicity. *A. racemosus* demonstrates neuroprotective potential against striatal neuronal damage as well as kainic acid induced hippocampal. The kainic acid (0.25 μ g/0.5 μ l) causes rise in protein carbonyl content and lipid peroxidation in anesthetized mice with decrease in the activities of glutathione and glutathione peroxidase. When *A. racemosus* extract was given to the mice then increase in the activity of glutathione peroxidase and reduction in glutathione contents, protein carbonyl contents and lipid peroxidation were observed [54]. There are reports that *A. racemosus* has non-selective inhibitory effects on acetylcholinesterase and butyrylcholinesterase owing to the existence of saponins

in methanolic extracts of roots relative to the chloroform and *n*-hexane fractions [55].

Antitussive Effect: Methanolic extracts of *A. racemosus* were investigated for their antitussive effects. Experiments were performed on albino mice having 5 groups, each group containing 10 mice. The control group was given 10ml/kg per oral dose of saline whereas 2nd and 3rd groups were provided with 200 and 400mg/kg per oral dose, respectively of *A. racemosus* extract. Codeine phosphate with oral doses of 10 and 20mg/kg were provided to 4th and 5th groups, respectively. Afterwards, cough was induced in all the mice by giving sulfur dioxide for one minute. The extracts displayed the dose dependent effects against sulfur dioxide induced cough; the doses of 200 and 400 mg/kg have caused 40 and 58.5% cough inhibition, respectively whereas codeine phosphate (standard antitussive drug) has displayed 36 and 55.4% inhibition of cough at 10 and 20 mg/kg, respectively. So it can be concluded that *A. racemosus* extracts can be used to treat cough [56].

Immunomodulatory Properties: *A. racemosus* has displayed significant effects on immune system. Investigations were done on bred male albino mice of 28 days age, by giving them oral doses of 1.5mg/kg of ochratoxin. Then the alcoholic extract of plant (100mg/kg) was fed to mice while control group was given only the distilled water. It resulted in the rise of tumor necrosis factor and IL-1 by macrophages in the mice (who received extract) relative to the control group [57]. Immunomodulatory effects were also exhibited in mice having tumor and taking cyclophosphamide. Cyclophosphamide causes a lowering in white cell total counts, the platelet counts and HA/HL antibody titers in mice whereas *A. racemosus* have enhances the antibody titer and white cell count [58]. The aqueous extract of *A. racemosus* roots has shown immunomodulatory effect in mice. It was observed that 100mg/kg oral dose of extract per body weight of mice had caused the modulation in Th1/Th2 immunity [59].

Antidepressant Effect: *A. racemosus* was examined for its antidepressant effects on rodents. The rodents received the methanolic extracts of *A. racemosus* roots standardized to saponins (62.2% w/w); the doses of 400, 200 and 100mg/kg were given for seven days on daily basis. Afterwards, the rodents were examined for antidepressant potential of plant extracts by performing learned helplessness test and forced swim test. The results have verified the antidepressant activity of *A. racemosus* which assists to the adrenergic and serotonergic systems [60]. Same kind of dose dependent antidepressant effects were also observed when methanolic extract (200, 100 and 50 mg/kg per oral dose) of *A. racemosus* was given to mice for 14 days [61].

Nootropic and Anti-amnesic Activity: Methanolic extract of *A. racemosus* root was tested for its memory enhancing (nootropic) and anti-amnesic potential. The male albino rats were subjected to 2 vital tests i.e., morris water maze (MWM) and elevated plusmaze (EPM) tests to test the memory/learning activity, respecti-

vely. Scopolamine and sodium nitrite-induced amnesic models were also used in rats to evaluate the anti-amnesic activity. The rats were pretreated with methanolic extract 50, 100 and 200mg/kg (p.o) of *A. racemosus* for seven days. Nootropic activity was shown by a considerable lowering of escape latency during MWM test. The scopolamine and sodium nitrite-induced rise in transfer latency was significantly reversed by MAR, thus demonstrating the anti-amnesic activity. Also, there was a dose-dependent inhibition of acetylcholinesterase enzyme in some particular regions of brain (hypothalamus, hippocampus and prefrontal cortex) [62].

Antiulcer Activity: *A. racemosus* and its phytoconstituents have been found effective in antiulcer therapy [63, 64]. Its root extracts have been reported to be diuretic tonic and demonstrate the ulcer healing effects probably *via* increasing the cytoprotection or mucosal resistance [13]. Its root extracts (methanolic) have displayed the ulcer protective effects and were used for the treatment of gastric ulcer in ancient Indian texts (Ayurvedic rasayana). The anti-ulcer effects of *A. racemosus* were investigated on gastroduodenal ulcer models by providing oral doses of 25-100 mg/Kg (2 times daily) for five days. There was significant decreases in duodenal ulcers (induced by cysteamineacute) and gastric ulcers (induced by aspirin plus pyloric ligation, pyloric ligation and cold restraint stress) at an oral dose of 50 mg/Kg (total saponins 0.9%) of *A. racemosus* root extracts. Also, after 10 days treatment, there was significant relief from chronic gastric ulcer induced by acetic acid. However, MAR have shown no effect on ethanol and aspirin-induced gastric ulcers. It was concluded that the mucosal defensive factors (like life span of cells, cellular mucus and mucus secretion) and anti-oxidant effect are significantly increased by MAR. However, MAR displays little or no effects on offensive factors like pepsin and acid [63].

Antiparasitic potential: In *vitro* anti-plasmodial and anti-leishmanial potential of water and methanolic extracts of *A. racemosus* were tested against *Leishmania major* and two laboratory-adapted *Plasmodium falciparum* isolates. The methanolic extracts have shown moderate anti-plasmodial activity with IC_{50} values of 33.95 and 32.63 μ g/ml against W2 and D6, respectively. However, the aqueous and methanolic extracts have shown leishmanicidal activity of $56.8 \pm 6.58\%$ and $58.3 \pm 8.22\%$, respectively. [65].

Antidiabetic Effect: From the ancient timings, numerous herbs had been employed for the treatment of diabetes. Their antidiabetic potential is owed to the presence of certain inorganic minerals (e.g., Ca, Zn, K and traces of Cr) which assist in the release of insulin from β -cells of islets of Langerhans and are helpful in the maintenance of normal glucose tolerance [66]. Earlier reports demonstrate the presence of numerous minerals in *A. racemosus* extract, out of which calcium is the most significant mineral for the insulin secretion. *A. racemosus* was reported for the lowering of blood pressure in rabbits and rats. The ethanol extract and ethyl acetate/chloroform/hexane partition fractions of its roots have shown concen-

tration dependent simulated insulin secretion in clonal β -cells, isolated rat islet cells and isolated perfused rat pancreas. The intracellular Ca^{+2} was increased by ethanol fraction of *A. racemosus* and its partition fractions with various solvents e.g., aqueous/butanol/ethyl acetate/chloroform/hexane [67].

Anticancer Activity: Many diosgenin and sarsapogenin derived steroids were separated from root extracts of *A. racemosus*. Its immunoside constituent has shown a valuable potential as an inducer of apoptosis of colon carcinoma cells [68]. The dried powdered extracts of *A. racemosus* were also reported for their cytotoxicity by brine shrimp inhibition [69]. There were investigations on rats having mammary tumor genesis induced by DMBA. The virgin female rats prior to their exposure to DBMA, were provided the diet of 2%, 1%, 0.5% and 0.25% root extract powder of *A. Asparagus*. It resulted in a sharp decline of mammary tumor incidence [70]. It was investigated in another study that shatavarin IV rich fraction (AR-2B) displays a valuable activity against cancer [71].

Aphrodisiac Activity: There is a lot of interest in the roots of *A. racemosus* for the treatment of sexual disorders. The aqueous and hydro-alcoholic extracts of *A. racemosus* roots were tested for their aphrodisiac activity. Experiments were done on male wistar albino rats with the extract doses of 400 and 200 mg/kg body weight. It was observed that hydroalcoholic extract was more active as compared to the aqueous extract in increasing the mount and mating performance in the experimental rats. [72].

Antibacterial Activity: *A. racemosus* roots were found to possess significant antibacterial potential [73, 74]. Its ethanolic extract (100, 300 and 500mg/ml) have shown considerable inhibition of *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Proteus mirabilis*, *Pseudomonas putida*, *Bacillus subtilis* and *Staphylococcus wernerii* by agar cup-plate method. The antibacterial activity was owed to the presence of various bioactive constituents including flavonoids, steroids, saponins, tannins, phenolic compounds, glycosides, carbohydrates and alkaloids in the root extract [73]. Significant *in vitro* antibacterial activity was also exhibited by methanolic extract (50, 100, 150 $\mu\text{g/mL}$) of *A. racemosus* roots against *S. aureus*, *B. subtilis*, *P. putida*, *S. typhimurium*, *S. typhi*, *V. cholerae*, *S.*, *S. sonnei*, *S. dysenteriae* and *E. coli* by disc diffusion method [75]. The biosynthesized copper nanoparticles in *A. racemosus* root have shown significant antimicrobial potential against two pathogens (*S. aureus* and *E. coli*) [76].

Anti-inflammatory Activity: The ethanolic extracts (200, 400, 600mg/kg) of *A. racemosus* leaves have displayed dose dependent anti-inflammatory effect in carrageenan induced paw oedema rats. By providing a dose concentration of 600mg/kg, 46 % inhibition was observed in experimental rats relative to the control group. This activity was owed to the presence of flavonoid and sterole type of compounds in *A. racemosus* leaf extract [77]. Methanolic extract of *A. racemosus* roots significantly inhibits the production of nitric oxide which shows a key role

in inflammation. It was concluded that the extract of *A. racemosus* can be employed for treatment of inflammation [78].

Antidiarrheal Effect: *A. racemosus* demonstrates a valuable potential against diarrhea [79]. The roots of *A. racemosus* is effective in the treatment of diarrhea as well as many skin diseases [80]. A significant antidiarrheal effect was observed when oral doses (200mg/kg) of aqueous or ethanolic extracts of *A. racemosus* were administered to the castor oil induced diarrheal rats (Albino Wistar). It also caused lowering of gastrointestinal motility in rats during charcoal meal test. It was concluded that *A. racemosus* extracts can be used as a herbal remedy for diarrhea treatment [81].

Increase of sperm count: The juice of *A. racemosus* (common name satamul) roots was traditionally used to increase the sperm count [82] and for the treatment of sterility issues of oligospermia. The lyophilized aqueous root extract has shown significant potential against oligospermia [83].

Antioxidant potential: Antioxidant are molecules that control and stop the oxidation by capturing the free radicals which cause the oxidation reactions [49]. The roots of *A. racemosus* had shown significant antioxidant potential during clinical trials in experimental animals [84]. The chloroform, *n*-hexane, petroleum ether, methanol, and ethanol extracts of *A. racemosus* roots had displayed *in vitro* antioxidant potential [85]. DPPH free radical scavenging assay had shown that ethanolic fraction possessed the highest IC₅₀ values (164.77 µg/ml) values as compared to the other fractions (methanol, vpet-ether, *n*-hexane and chloroform). It is due to the fact that ethanolic fraction was rich in total phenolic contents (108.78 mg/gm, Gallic Acid Equivalent) as compared to other fractions. This study clarified that *A. racemosus* may be a source of valuable antioxidant ingredients [49]. The antioxidant potential of *A. racemosus* methanolic extract was observed to be highest (21.99 g GAE/L), whereas the antioxidant activity of an *n*-hexane extract was lowest (5.87 g GAE/L). Ethyl acetate (13.13 AE/L) and *n*-hexane (3.92 AE/L) extracts were found to have the highest and lowest levels, respectively of alpha-amylase inhibition. The presence of stigmasterol was identified as the cause of the amylase inhibition in the deep blue zone of the hRF = 72 extracts on the TLC plate [86]. The extraction of *A. racemosus* Willd roots was made by employing the saturated CO₂ fluid technique and ethanol. The polyphenolic components like quercetin, naringenin and *p*-coumaric acid were found abundant in the extract. Its DPPH scavenging potential can be comparable to that of vitamin C [87].

Hepatoprotective Effect: The hepatoprotective effects of *A. racemosus* were evaluated by inducing malignancy in Wistar rats by treating them with diethylnitrosamine (200mg/kg b wt, i.p.) once a week for two weeks. The rats were then treated with DDT (0.05% in diet) which is a tumor promoter. The p53+ foci were found in hepatic tissues of rats treated with diethylnitrosamine. However, when the hepatic tissues of Wistar rats were pretreated with the aqueous root extracts of

A. Racemosus, the incidence of hepatocarcinogenesis was not happened and the development of p53+ foci was also not observed. The results have shown that *A. Racemosus* extract is responsible for reforming the hepatotoxicity and oxidative stress caused by DEN treatment. It was concluded that aqueous root extract of *A. racemosus* acts as an efficient formulation for the prevention of hepatocarcinogenesis caused by DEN treatment [50]. *A. racemosus* also displayed hepatoprotective activities against the paracetamol induced hepatotoxicity in rats. A dose of 100 mg/kg b. wt. caused significant alteration of antioxidant status and serum marker enzymes to a normal level in experimental rats. The observed activities were comparable to silymarin (100 mg/kg b. wt. p.o.) [88].

Hypocholestrimic Effect: Hypocholestrimic effect was observed in streptozotocin-induced diabetic rats. The doses (200 and 400 mg/kg/b.w) of *A. Racemosus* (Wild) ethanolic extract to rats for 21 days have caused a significant lowering of the cholesterol and triglyceride levels in rats. After a dose treatment of 200 and 400mg/kg/b.w., the original cholesterol level (130.9 mg/dl) in rats was decreased to 102.0 and 87.68 mg/dl, respectively whereas triglyceride level was reduced from 143.8 to 27.60 and 22.33 mg/dl, respectively. It was demonstrated that *A. racemosus* has a good hypocholesterolemic effect [89]. In another investigation, the root fraction (5 and 10 g per doses for 4 weeks) of *A. racemosus* was introduced in hypercholesteremic rats. This treatment had resulted in a decrease of cholesterol, triglycerides, VLDL and LDL levels and rise in fecal secretion of blood cholesterol, sterol and bile acid. It also caused an increase of hepatic HMG-CoA reductase activity and bile acid content [90].

Hydroalcoholic and hexane extracts of *A. Racemosus* were tested by disc diffusion method for antifungal potential against *Candida Albicans* and *Aspergillus Niger*. They have shown the antifungal potential against *A. Niger*, with greater activity displayed by hydroalcoholic extract. However, no activity was displayed against *C. Albicans*. The hydroalcoholic and hexane fraction have shown zones of inhibitions of 20 and 10 mm, respectively against *A. niger* [91]. The methanolic extracts of *A. racemosus* roots and tubers have also shown *in vitro* antifungal potential against *Candida stellatoidea*, *Candida parapsilosis*, *Candida guilliermondii*, *Candida krusei*, *Candida tropicalis* and *Candida albicans* by disc diffusion method. So it was found that root and tubers extracts of *A. Racemosus* can be utilized to treat vaginal candidiasis [92].

Wound Healing Potential: The wound healing potential (*in vivo*) of aqueous extract of *A. racemosus* roots was observed on albino rats by using excision and incision wound model. The oral doses of 400mg/kg and 200mg/kg were administered for 10 to 22 days. It resulted in an increased skin breaking strength, significant increase of wound contraction rate and remarkable improvement in epithelialisation period. The results have suggested the probable use of this plant in wound healing [93].

For Gynecological Disorders: Ethno's medicinal study have shown that root paste of *A. racemosus* can be directly utilized on the abdomen for assistance in delivery [94]. A survey had shown that *A. racemosus* is used to treat a number of problems including menorrhagia, leucorrhoea, dryness of sexual organs, miscarriages and inflammatory conditions of sexual organs. The plant extract also boosts libido, augments ovulation and folliculogenesis, normalizes the uterus (called as a postpartum tonic) and also prepares the womb for conception [11].

Anti-urolithiatic Activity: The ethanolic root extracts of *A. racemosus* have shown anti-urolithiatic activity which was comparable to that of cystone (Shashi et al, 2009). The ethanolic extracts of *A. racemosus* were tested against urolithiasis in rats. For this purpose, six groups (n = 6) were made of 36 male Wistar albino rats on a random basis. One group (Group I) was selected as a control group whereas all the other groups (Groups II-VI) were fed for ten days with ethylene glycol (0.75%) and ammonium chloride (2%) to induce urolithiasis. The ethanolic extract of *A. racemosus* was also fed to the rates for ten days at doses of 200 mg/kg (groups III), 400 mg/kg (groups IV), 800 mg/kg (groups V) and 1600 mg/kg (groups VI). Group II (positive control) rats were given only EG/AC. The distilled water (6 µl/g) and drinking water were administered by gavage to the rates of group I. The blood samples were examined after ten days. The kidneys were removed, sectioned and their histopathological examination was also performed. It was found that serum concentrations of creatinine, urea, phosphorus, and calcium were lowered in rats receiving doses of 800 and 1600 mg/kg doses of ethanolic extracts of *A. racemosus*; also, both these groups (V and VI) have shown less tissue damage and their kidneys were almost similar to group I rats. It was concluded that ethanolic extracts of *A. racemosus* can be used to prevent urolithiasis [95].

Anti-sebum Activity: Overactive sebaceous glands that produce oily skin have an impact on personality and self-esteem. The expression of the SRD5A gene, which codes for steroid 5-alpha reductase, has been linked to the secretion of face sebum. *A. racemosus* wild root extract has been found to affect on SRD5A mRNA expression regulation and anti-sebum potency in male volunteers. The extract caused significant improvement in facial secretion with the lowering in the percentages of pore area after treatment of 15 and 30 days. The mRNA expression of SRD5A1 and SRD5A2 was reduced to about 45.45% and 90.86%, respectively by root extract [87].

Traditional Uses: *A. racemosus* is well-known for treating numerous diseases including urinary tract infections, leucorrhoea, benign prostatic hyperplasia, anxiety disorders, dysmenorrhea, angina, hypertension and hyperlipidemia etc. This plant contains a variety of secondary metabolites but its main contents are steroidal saponins, which are responsible for numerous biological actions [10]. Oral administration of root powder of *A. racemosus* along with cow's butter-milk for 3-4 days has been found effective for kidney stones [96]. *A. racemosus* is also used to treat skin problems (mixture of *A. racemosus* tubers and *Plumbago indica*

leaves) [92, 94], lactation issues (root powder along with milk or honey enhances the amount of milk) and leucorrhoea (root decoction) [97]. There are reports that root extract of *A. racemosus* is effective in the treatment of dysentery, diarrhoea, and general weakness [38, 98]. The tuberous root of *A. racemosus* was traditionally used to treat epilepsy by taking for 90 days once a day ½ cup decoction with milk [99]. The use of root juice for rise of sperm count and for impeding blood dysentery has also been reported [82].

Side Effects: Teratological disorders were observed when 1x10⁵ug/kg/day extract of *A. racemosus* was given to the pregnant rats for 60 days. In test groups, gross malformations and an enhanced resorption of fetuses were observed e.g., intrauterine growth retardation with a small placental size and swelling in legs. The live pup displayed a considerable delay in different developmental factors and a decline in body length and body weight [100]. *A. racemosus* has a minor diuretic effect, positive chronotropic and ionotropic effects, and causes hypotension in cats. It showed a depression of respiration in cats, decrease of blood flow in the mesenteric vessels of mice and bronchial muscle dilation in pigs [101, 102].

Importance in Drugs: *A. racemosus* is a part of many important drugs which are summarized in **table 1**, along with their contents and medicinal uses.

TABLE 1 Various formulations of *A. Racemosus*

SR. NO.	DRUG	CONTENT OF <i>A. RACEMOSUS</i>	MEDICINAL PROPERTIES/USES	REFERENCE
1	Abana	10 mg Shatavari root extract per tablet	Mild to moderate hypertension, Hyperlipidemic conditions	[103] [104]
2	Diabecon	20 mg Shatavari root extract per tablet	Monotherapy in non-insulin-dependent diabetes mellitus	[28, 105, 106]
3	EveCare	32 mg Shatavari root extract per 5 ml syrup	Metrorrhagia Oligomenorrhoea Dysmenorrhoea Menorrhagia	[28]
4	Geriforte	20 mg Shatavari root powder per tablet	Stress related anxiety Prolonged illness and convalescence, Generalized anxiety disorders, Geriatric stress	[107]
5	Himplasia	80 mg Shatavari root powder per tablet	Benign prostatic hyperplasia	[108]
6	Lukol	40 mg Satavari root extract per tablet	Pelvic inflammatory disease, Malaise Backache associated with leukorrhoea and Leukorrhoea	[109]
7	Renalka	50mg shatavari root extract per 5mL of syrup	Hematuria, Dysuria, Recurrent Urinary Tract Infection, Burning micturition Cystitis,	[110]

MEDICINAL VALUE

Plants are commonly rich in nutritional contents and important for diet [111-113]. The food application and health benefits of *A. racemosus* powder were investigated by numerous researchers [114, 115]. Milk secretion lactating females is enhanced when they use Shatavari fortified milk and milk products. Some bakery products can also be fortified with Shatavari extracts to increase the nutritional characteristics of these food items [116]. *A. racemosus* is rich in many nutrients and may act as an important component of feed supplements in the animal diets. The analysis of its crude fiber, crude protein, ether extract, ash content and nitrogen free extract have shown that this herb is very rich in nitrogen free extract and minerals including Zn, Cu, Fe, Mg, Ca etc. This plant has positive effects on reproduction capacity and milk production of dairy animals. It can lower the stress of dairy animals and increase the productivity of clean and healthy milk. In cows, it can prevent the infection of the udder and reproductive organs by boosting the immune system [117]. It has been reported that supplementation of ½ KG fresh roots of *A. shatavari* on daily basis at the time of milking in buffaloes increases significant yield of milk [118]. A similar outcome was also observed in freshly calved crossbred cattle, where oral supplementation (100g per animal) with root powder of *A. shatavari* on alternate days significantly increased milk production [119]. It is believed that feeding dairy cows with *A. racemosus* root powder supplements at various lactation stages enhanced their reproductive efficiency and nutritional characteristics [120].

CONCLUSIONS

Asparagus racemosus (shatavari) contains numerous steroidal saponins, steroids, alkaloids, flavonoids, cyclic hydrocarbons, tannins, anthraquinones, polysaccharides, oligospirostanoside, trace minerals, derivatives of dihydrophenanthrene and derivatives of furans and essential oils. *A. racemosus* has shown neuroprotective, antitussive, immunomodulatory, antidepressant, nootropic, anti-amnesic, anti-ulcer, antiparasitic, antidiabetic, anticancer, aphrodisiac, antibacterial, anti-inflammatory, antidiarrheal, antioxidant, hepatoprotective, hypocholesterimic, anti-rolithiatic, anti-sebum and wound healing effects. It has been found effective in improving sperm count and also used to treat blood dysentery, skin problems, kidney stones, dysentery, diarrhoea, epilepsy, general weakness, leucorrhoea, gynecological disorders, lactation issues and teratological disorders. Its root extract is an important ingredient of many useful drug formulations including Abana, Diabecon, EyeCare, Geriforte, Himplasia, Lukol and Renalka. Its roots are rich in many nutrients and can be orally given to dairy animals as food supplements to increase the milk production, reproduction capacity and immune system.

REFERENCES

- [1] Hussain S, Javed M, Abid MA, Khan MA, Syed SK, Faizan M, Feroz F. *Prunus Avium L.*; Phytochemistry, Nutritional and Pharmacological Review. *Advancements in Life Sciences*. 2021;8(4):307-14.
- [2] Butt SZ, Hussain S, Munawar KS, Tajammal A, Muazzam MA. Phytochemistry of *Ziziphus Mauritiana*; its Nutritional and Pharmaceutical Potential. *Scientific Inquiry and Review*. 2021;5(2):1-15.
- [3] Hussain S, Zamir I, Javed M, Munawar KS, Batool I. Phytochemical Composition of Ginger, its Nutritional and Pharmacological Importance. *Lahore Garrison University Journal of Life Sciences*. 2020;4(01):17-31.
- [4] Naseer S, Hussain S, Naeem N, Pervaiz M, Rahman M. The phytochemistry and medicinal value of *Psidium guajava* (guava). *Clinical Phytoscience*. 2018;4(1):1-8.
- [5] Amjad M, Hussain S, Javaid K, Khan A, Ali B, Noreen M, Khan AR, Hussain G, Ullah H. Plant representation, phytochemistry and medicinal assets of *Asparagus racemosus*. *The Pharmaceutical and Chemical Journal*. 2020;7(5):12-23.
- [6] Saeed M, Naseer S, Hussain S, Iqbal M. Phytochemical composition and pharmacological effects of *Cassia fistula*. *Scientific Inquiry and Review*. 2020;4(1):59-69.
- [7] Naseer S, Hussain S, Zahid Z. Nutritional and antioxidant potential of common vegetables in Pakistan. *RADS Journal of Biological Research and Applied Sciences*. 2019;10(1):36-40.
- [8] Riaz M, Fatima H, ur Rehman MM, Qadir R, Hussain S, Hafeez A, Siddique AB. Appraisal of antioxidant potential and biological studies of bogan bail (*Bougainvillea glabra*) leaf extracts using different solvents. *Czech Journal of Food Sciences*. 2021;39(3):176-80.
- [9] Naseer S, Afzal M, Nisa A, Hussain S, Ahmad M, Parveen S, Anjum F, Riaz M. Extraction of brown dye from *Eucalyptus* bark and its applications in food storage. *Quality Assurance and Safety of Crops and Foods*. 2019;11(8):769-80.
- [10] Ganapathy AA, Priya VH, Kumaran A. Medicinal plants as a potential source of Phosphodiesterase-5 inhibitors: A review. *Journal of Ethnopharmacology*. 2021;267:113536.
- [11] Sharma K. *Asparagus racemosus* (Shatavari): A versatile female tonic. *International Journal of Pharmaceutical and Biological Archive*. 2011;2(3).
- [12] Bopana N, Saxena S. *Asparagus racemosus*--Ethnopharmacological evaluation and conservation needs. *Journal of ethnopharmacology*. 2007;110(1):1-15.
- [13] Alok S, Jain SK, Verma A, Kumar M, Mahor A, Sabharwal M. Plant profile, phytochemistry and pharmacology of *Asparagus racemosus* (Shatavari): A review. *Asian Pacific journal of tropical disease*. 2013;3(3):242-51.
- [14] Shodhini. *Touch Me, Touch-me-not: Women, Plants, and Healing: Kali for Women*; 1997.
- [15] Simon D. *Wisdom of Healing*. Random House Value Publishing; 1999.
- [16] Ali MS, Mukherjee S, Roy D, Pal G, Makar S. *Asparagus Racemosus*, a Climbing Ayurvedic Medicinal Plant: Review on its Cultivation, Morphology and Medicinal Significance. *PharmaTutor*. 2018;6(12):46-54.
- [17] Thakur S, Kaurav H, Chaudhary G. *Shatavari (Asparagus Racemosus)-The Best Female Reproductive Tonic*. 2021;8(5): 73-84.
- [18] Kumeta Y, Maruyama T, Wakana D, Kamakura H, Goda Y. Chemical analysis reveals the botanical origin of shatavari products and confirms the absence of alkaloid asparagamine A in *Asparagus racemosus*. *Journal of natural medicines*. 2013;67(1):168-73.
- [19] Hayes PY, Jahidin AH, Lehmann R, Penman K, Kitching W, De Voss JJ. Steroidal saponins from the roots of *Asparagus racemosus*. *Phytochemistry*. 2008;69(3):796-804.
- [20] Sharma P, Chauhan PS, Dutt P, Amina M, Suri KA, Gupta BD, Suri OP, Dhar KL, Sharma D, Gupta V. A unique immuno-stimulant steroidal saponin acid from the roots of *Asparagus racemosus*. *Steroids*. 2011;76(4):358-64.
- [21] Ahmad S, Ahmad S, Jain P. Chemical examination of Shatavari (*Asparagus racemosus*). *Bull Medico Ethnobotanical Res*. 1991;12(3-4):4.
- [22] Khare CP. *Indian medicinal plants: an illustrated dictionary*: Springer Science and Business Media; 2008.

- [23] Paliwal M, Siddiqui I, Singh J, Tiwari H. Chemical examination of roots of *Asparagus racemosus*. *Journal of the Indian Chemical Society*. 1991;68(7):427-8.
- [24] Bose S, Show S, Hazra M, Sarkar T. Comparative study of Antioxidant Activity of Herbal Drugs and their Formulations using *Asparagus racemosus* and *Centella asiatica*. *American Journal of PharmTech Research*. 2012;2:391-8.
- [25] Sabde S, Bodiwala HS, Karmase A, Deshpande PJ, Kaur A, Ahmed N, Chauthe SK, Brahmabhatt KG, Phadke RU, Mitra D. Anti-HIV activity of Indian medicinal plants. *Journal of natural medicines*. 2011;65(3):662-9.
- [26] Sharma U, Kumar N, Singh B, Munshi RK, Bhalerao S. Immunomodulatory active steroidal saponins from *Asparagus racemosus*. *Medicinal Chemistry Research*. 2013;22(2):573-9.
- [27] Sekine T, Ikegami F, Fukasawa N, Kashiwagi Y, Aizawa T, Fujii Y, Ruangrungsi N, Murakoshi I. Structure and relative stereochemistry of a new polycyclic alkaloid, asparagamine A, showing anti-oxytocin activity, isolated from *Asparagus racemosus*. *Journal of the Chemical Society, Perkin Transactions I*. 1995(4):391-3.
- [28] Singla R, Jaitak V. Shatavari (*Asparagus Racemosus* Wild): A review on its cultivation, morphology, phytochemistry and pharmacological importance. *International Journal of Pharmacy and Life Sciences*. 2014;5(3).
- [29] Saxena V, Chourasia S. A new isoflavone from the roots of *Asparagus racemosus*. *Fitoterapia*. 2001;72(3):307-9.
- [30] Bopana N, Saxena S. *Asparagus racemosus*—Ethnopharmacological evaluation and conservation needs. *Journal of ethnopharmacology*. 2007;110(1):1-15.
- [31] Mandal D, Banerjee S, Mondal NB, Chakravarty AK, Sahu NP. Steroidal saponins from the fruits of *Asparagus racemosus*. *Phytochemistry*. 2006;67(13):1316-21.
- [32] Sekine T, Fukasawa N, Murakoshi I, Ruangrungsi N. A 9, 10-dihydrophenanthrene from *Asparagus racemosus*. *Phytochemistry*. 1997;44(4):763-4.
- [33] Wiboonpun N, Phuwapraisirisan P, Tip-pyang S. Identification of antioxidant compound from *Asparagus racemosus*. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*. 2004;18(9):771-3.
- [34] Hayes PY, Jahidin AH, Lehmann R, Penman K, Kitching W, De Voss JJ. Asparinins, asparosides, curillins, curilloides and shavatarins: structural clarification with the isolation of shatavarin V, a new steroidal saponin from the root of *Asparagus racemosus*. *Tetrahedron letters*. 2006;47(49):8683-7.
- [35] Joshi J, Dev S. Chemistry of Ayurvedic Crude Drugs. Part 8. Shatavari-2: Structure Elucidation of Bioactive Shatavarin-I and Other Glycosides. *ChemInform*. 1988;19(28).
- [36] Sekine T, Ikegami F, Fukasawa N, Kashiwagi Y, Aizawa T, Fujii Y, Ruangrungsi N, Murakoshi I. Structure and relative stereochemistry of a new polycyclic alkaloid, asparagamine A, showing anti-oxytocin activity, isolated from *Asparagus racemosus*. *Journal of the Chemical Society, Perkin Transactions I*. 1995;1(4):391-3.
- [37] Acharya S, Acharya N, Bhangale J, Shah S, Pandya S. Antioxidant and hepatoprotective action of *Asparagus racemosus* Willd. root extracts. *Indian Journal of Experimental Biology*. 2012;50(11):795–801.
- [38] Hasan N, Ahmad N, Zohrameena S, Khalid M, Akhtar J. *Asparagus racemosus*: for medicinal uses and pharmacological actions. *International Journal of Advanced Research*. 2016;4(3):259-67.
- [39] Tambvekar N. Ayurvedic drugs in common eye conditions. *Journal of the National Integrated Medical Association*. 1985;27(5):13-8.
- [40] Mohanta B, Chakraborty A, Sudarshan M, Dutta RK, Baruah M. Elemental profile in some common medicinal plants of India. Its correlation with traditional therapeutic usage. *Journal of radio-analytical and nuclear chemistry*. 2003;258(1):175-9.
- [41] Sharma S. Constituents of the fruits of *asparagus racemosus* willd. *Pharmazie*. 1981;36(10):709.
- [42] Choudhary B, Kar A. Mineral contents of *Asparagus racemosus*. *Indian Drugs*. 1992;29(13):623.
- [43] Saxena V, Chourasia S. A new isoflavone from the roots of *Asparagus racemosus*. *Fitoterapia*. 2001;72(3):307-9.

- [44] Sekine T, Fukasawa N, Murakoshi I, Ruangrunsi N. A 9, 10-dihydrophenanthrene from *Asparagus racemosus*. *Phytochemistry*. 1997;44(4):763-4.
- [45] Chawla A, Chawla P, Mangalesh R. *Asparagus racemosus* (Willd): Biological Activities and its Active Principles. *Indo Global Journal of Pharmaceutical Sciences*. 2011;2:113-20.
- [46] Mandal D, Banerjee S, Mondal NB, Chakravarty AK, Sahu NP. Steroidal saponins from the fruits of *Asparagus racemosus*. *Phytochemistry*. 2006;67(13):1316-21.
- [47] Hayes PY, Jahidin AH, Lehmann R, Penman K, Kitching W, De Voss JJ. Asparinins, asparosides, curillins, curillosides and shavatarins: structural clarification with the isolation of shatavarin V, a new steroidal saponin from the root of *Asparagus racemosus*. *Tetrahedron letters*. 2006;47(49):8683-7.
- [48] Sidiq T, Khajuria A, Suden P, Singh S, Satti N, Suri K, Srinivas V, Krishna E, Johri R. A novel sarsasapogenin glycoside from *Asparagus racemosus* elicits protective immune responses against HBsAg. *Immunology letters*. 2011;135(1):129-35.
- [49] Hossain MI, Sharmin FA, Akhter S, Bhuiyan MA, Shahriar M. Investigation of cytotoxicity and in-vitro antioxidant activity of *Asparagus racemosus* root extract. *International Current Pharmaceutical Journal*. 2012;1(9):250-7.
- [50] Agrawal A, Sharma M, Rai SK, Singh B, Tiwari M, Chandra R. The effect of the aqueous extract of the roots of *Asparagus racemosus* on hepatocarcinogenesis initiated by diethylnitrosamine. *Phytotherapy Research*. 2008;22(9):1175-82.
- [51] Sharma U, Saini R, Kumar N, Singh B. Steroidal saponins from *Asparagus racemosus*. *Chemical and Pharmaceutical Bulletin*. 2009;57(8):890-3.
- [52] Wiboonpun N, Phuwapraisrisan P, Tip-pyang S. Identification of antioxidant compound from *Asparagus racemosus*. *Phytotherapy Research*. 2004;18(9):771-3.
- [53] Sharma A, Sahrawat A. *Asparagus racemosus*-wonder plant. *International Journal*. 2014;2(4):1039-45.
- [54] Parihar M, Hemnani T. Experimental excitotoxicity provokes oxidative damage in mice brain and attenuation by extract of *Asparagus racemosus*. *Journal of neural transmission*. 2004;111(1):1-12.
- [55] Meena J, Ojha R, Muruganandam A, Krishnamurthy S. *Asparagus racemosus* competitively inhibits in vitro the acetylcholine and monoamine metabolizing enzymes. *Neuroscience letters*. 2011;503(1):6-9.
- [56] Mandal SC, Kumar CK A, Mohana Lakshmi S, Sinha S, Murugesan T, Saha B, Pal M. Antitussive effect of *Asparagus racemosus* root against sulfur dioxide-induced cough in mice. *Fitoterapia*. 2000;71(6):686-9.
- [57] Dhuley J. Effect of some Indian herbs on macrophage functions in ochratoxin A treated mice. *Journal of ethnopharmacology*. 1997;58(1):15-20.
- [58] Diwanay S, Chitre D, Patwardhan B. Immunoprotection by botanical drugs in cancer chemotherapy. *Journal of ethnopharmacology*. 2004;90(1):49-55.
- [59] Gautam M, Saha S, Bani S, Kaul A, Mishra S, Patil D, Satti N, Suri K, Gairola S, Suresh K. Immunomodulatory activity of *Asparagus racemosus* on systemic Th1/Th2 immunity: Implications for immunoadjuvant potential. *Journal of ethnopharmacology*. 2009;121(2):241-7.
- [60] Singh GK, Garabadu D, Muruganandam A, Joshi VK, Krishnamurthy S. Antidepressant activity of *Asparagus racemosus* in rodent models. *Pharmacology Biochemistry and Behavior*. 2009;91(3):283-90.
- [61] Dhingra D, Kumar V. Pharmacological evaluation for antidepressant like activity of *Asparagus racemosus* wild in mice. *Pharmacologyonline*. 2007;3:133-52.
- [62] Ojha R, Sahu AN, Muruganandam A, Singh GK, Krishnamurthy S. *Asparagus racemosus* enhances memory and protects against amnesia in rodent models. *Brain and cognition*. 2010;74(1):1-9.
- [63] Sairam K, Priyambada S, Aryya N, Goel R. Gastroduodenal ulcer protective activity of *Asparagus racemosus*: an experimental, biochemical and histological study. *Journal of ethnopharmacology*. 2003;86(1):1-10.
- [64] Sen S, Chakraborty R, De B, Mazumder J. Plants and phytochemicals for peptic ulcer: An overview. *Pharmacognosy reviews*. 2009;3(6):270.
- [65] Kigundu EV, Rukunga GM, Keriko JM, Tonui WK, Gathirwa JW, Kirira PG, Irungu B, Ingonga JM, Ndiege IO. Anti-parasitic activity and cytotoxicity of selected medicinal plants from Kenya. *Journal of ethnopharmacology*. 2009;123(3):504-9.

- [66] Kar A, Choudhary B, Bandyopadhyay N. Preliminary studies on the inorganic constituents of some indigenous hypoglycaemic herbs on oral glucose tolerance test. *Journal of ethnopharmacology*. 1999;64(2):179-84.
- [67] Hannan J, Marenah L, Ali L, Rokeya B, Flatt PR, Abdel-Wahab YH. Insulin secretory actions of extracts of *Asparagus racemosus* root in perfused pancreas, isolated islets and clonal pancreatic beta-cells. *Journal of endocrinology*. 2007;192(1):159-68.
- [68] Bhutani K, Paul A, Fayad W, Linder S. Apoptosis inducing activity of steroidal constituents from *Solanum xanthocarpum* and *Asparagus racemosus*. *Phytomedicine*. 2010;17(10):789-93.
- [69] Potduang B, Meeploy M, Giwanon R, Benmart Y, Kaewduang M, Supatanakul W. Biological activities of *Asparagus racemosus*. *African Journal of Traditional, Complementary and Alternative Medicines*. 2008;5(3):230-7.
- [70] Rao A. Inhibitory action of *Asparagus racemosus* on DMBA induced mammary carcinogenesis in rats. *International journal of cancer*. 1981;28(5):607-10.
- [71] Mitra SK, Prakash NS, Sundaram R. Shatavarins (containing Shatavarin IV) with anticancer activity from the roots of *Asparagus racemosus*. *Indian journal of pharmacology*. 2012;44(6):732.
- [72] Wani JA, Achur RN, Nema R. Phytochemical screening and aphrodisiac activity of *Asparagus racemosus*. *International Journal of Pharmaceutical Sciences and Drug Research*. 2011;8:9.
- [73] Ravishankar K, Kiranmayi G, Lalitha TM, Priyanka T, Ranjith T, Someswarao S, Raju VK, Divya A. Preliminary phytochemical screening and in vitro antibacterial activity on *Asparagus racemosus* root extract. *International Journal of Pharmaceutical, Chemical and Biological Sciences*. 2012;2:117-23.
- [74] Umashanker M, Shruti S. Traditional Indian herbal medicine used as antipyretic, antiulcer, anti-diabetic and anticancer: A review. *International Journal of Research in Pharmacy and Chemistry*. 2011;1(4):1152-9.
- [75] Mandal SC, Nandy A, Pal M, Saha B. Evaluation of antibacterial activity of *Asparagus racemosus* Willd. root. *Phytotherapy Research*. 2000;14(2):118-9.
- [76] Subha V, Thulasimuthu E, Ilangoan R. Bactericidal action of copper nanoparticles synthesized from methanolic root extract of *Asparagus racemosus*. *Materials Today: Proceedings*. 2022;64:1761-7.
- [77] Battu G, Kumar B. Anti-inflammatory activity of leaf extract of *Asparagus racemosus* Willd. *International Journal of Chemical Sciences*. 2010;8(2):1329-38.
- [78] Kanwar AS, Bhutani KK. Effects of *Chlorophytum arundinaceum*, *Asparagus adscendens* and *Asparagus racemosus* on pro-inflammatory cytokine and corticosterone levels produced by stress. *Phytotherapy Research*. 2010;24(10):1562-6.
- [79] Gomase V, Sherkhane A. Isolation, structure elucidation and biotransformation studies on secondary metabolites from *Asparagus racemosus*. *International Journal of Microbiology Research*. 2010;2:1.
- [80] Parihar S, Sharma D. A brief overview on *Asparagus racemosus*. *IJRAR*. 2021;8(4):96-108.
- [81] Venkatesan N, Thiyagarajan V, Narayanan S, Arul A, Raja S, Kumar SV, Rajarajan T, Perianayagam JB. Anti-diarrhoeal potential of *Asparagus racemosus* wild root extracts in laboratory animals. *J Pharm Pharmaceut Sci*. 2005;8(1):39-46.
- [82] Anisuzzaman M, Rahman A, Harun-Or-Rashid M, Naderuzzaman A, Islam A. An ethnobotanical study of Madhupur, Tangail. *Journal of Applied Sciences Research*. 2007;3(7):519-30.
- [83] Thakur M, Dixit V. Effect of some vajikaran herbs on peniculation activities and in vitro sperm count in male. *Sexuality and Disability*. 2007;25(4):203-7.
- [84] Pandey AK, Gupta A, Tiwari M, Prasad S, Pandey AN, Yadav PK, Sharma A, Sahu K, Asrafuzman S, Vengayil DT. Impact of stress on female reproductive health disorders: Possible beneficial effects of shatavari (*Asparagus racemosus*). *Biomedicine and Pharmacotherapy*. 2018;103:46-9.
- [85] Karuna D, Dey P, Das S, Kundu A, Bhakta T. In vitro antioxidant activities of root extract of *Asparagus racemosus* Linn. *Journal of traditional and complementary medicine*. 2018;8(1):60-5.
- [86] Das P, Ashraf GJ, Baishya T, Dua TK, Paul P, Nandi G, Sahu R. High-performance thin-layer chromatography coupled attenuated total reflectance-Fourier-transform infrared and NMR spectroscopy-based identification of α -amylase inhibitor from the aerial part of *Asparagus racemosus* Willd. *Phytochemical Analysis*. 2022.

- [87] Ruksiriwanich W, Khantham C, Linsaenkart P, Chaitep T, Jantrawut P, Chittasupho C, Rachtanapun P, Jantanasakulwong K, Phimolsiripol Y, Sommano SR. In vitro and in vivo regulation of SRD5A mRNA expression of supercritical carbon dioxide extract from *Asparagus racemosus* Willd. Root as anti-sebum and pore-minimizing active ingredients. *Molecules*. 2022;27(5):1535.
- [88] El-Sensiy YA, Ahmad SA, Farid AS. Hepatoprotective effect of *asparagus racemosus* in paracetamol induced hepatotoxicity in rats. *Benha Veterinary Medical Journal*. 2015;28(1):133-7.
- [89] Vadivelan R, Dipanjan M, Umasankar P, Dhanabal SP, Satishkumar MN, Antony S, Elango K. Hypoglycemic, antioxidant and hypolipidemic activity of *Asparagus racemosus* on streptozotocin-induced diabetic in rats. *Advances in Applied Science Research*. 2011;2(3):179-85.
- [90] Bharati J, Kumar S. *Shatavari (Asparagus racemosus)*. Chapter 29. *Phytobiotics and Animal Production*. 567-590.
- [91] Mathur A, Singh R, Yousuf S, Bhardwaj A, Verma SK, Babu P, Gupta V, Prasad G, Dua V. Antifungal activity of some plant extracts against Clinical Pathogens. *Adv Appl Sci Res*. 2011;2:260-4.
- [92] Singh R, Geetanjali. *Asparagus racemosus*: a review on its phytochemical and therapeutic potential. *Natural Product Research*. 2016;30(17):1896-908.
- [93] Kumar S, Rajput R, Patil V, Udupa A, Gupta S, Rathnakar U, Rao S, Benegal D, Benegal A, Shubha H. Wound healing profile of *Asparagus racemosus* (Liliaceae) Wild. *Current Pharma Research*. 2011;1(2):111-4.
- [94] Shaha P, Bellankimath A. Pharmacological profile of *Asparagus racemosus*: A review. *International Journal of Current Microbiology and Applied Sciences*. 2017;6(11):1215-23.
- [95] Jagannath N, Chikkannasetty SS, Govindadas D, Devasankaraiah G. Study of antiulcer activity of *Asparagus racemosus* on albino rats. *Indian Journal of Pharmacology*. 2012;44(5):576.
- [96] Rajakumar N, Shivanna M. Traditional herbal medicinal knowledge in Sagar taluk of Shimoga district, Karnataka, India. *Indian Journal of Natural Products and Resources*. 2010;1(1):102-8.
- [97] Yadav J, Kumar S, Siwach P. Folk medicine used in gynecological and other related problems by rural population of Haryana. *Indian Journal of Traditional Knowledge*. 2006;5(3):323-6.
- [98] Senthilkumar M, Gurumoorthis P, Janardhanan K. Some medicinal plants used by Irular, the tribal people of Marudhamalai hills, Coimbatore, Tamil Nadu. *Natural Product Radiance*. 2006;5(5):382-8.
- [99] Das HB, Majumdar K, Datta B, Ray D. Ethnobotanical uses of some plants by Tripuri and Reang tribes of Tripura. *Natural Product Radiance*. 2009;8(2):172-80.
- [100] Gutiérrez-Rebolledo GA, Drier-Jonas S, Jiménez-Arellanes MA. Natural compounds and extracts from Mexican medicinal plants with anti-leishmaniasis activity: An update. *Asian Pacific Journal of Tropical Medicine*. 2017;10(12):1105-10.
- [101] Kumar MS, Udupa A, Sammodavardhana K, Rathnakar U, Shvetha U, Kodancha G. Acute toxicity and diuretic studies of the roots of *Asparagus racemosus* willd in rats. *West Indies medical journal*. 2010;59(1):3-5.
- [102] Goyal R, Singh J, Lal H. *Asparagus racemosus*--an update. *Indian Journal of Medical Sciences*. 2003;57(9):408.
- [103] Venkataramaiah H. Double-blind comparative clinical trial of Abana and Simvastatin in Hyperlipidaemia. *Insertion in Stroke* Feb-Mar. 2002;1001.
- [104] [104] Dubey G, Agrawal A, Srivastava Sr V, Agrawal U, Udupa K. Management of Risk Factors of Coronary Heart Disease with an Indigenous Compound--Abana (A Controlled Study). *Probe*. 1985;25:1-46.
- [105] Kohli K, Shilin G, Kolhapure S. Evaluation of the clinical efficacy and safety of Diabecon in NIDDM. *The Antiseptic*. 2004;101(11):487-94.
- [106] Seshaiya V, Agrawal J, Maji D, Yajnik V, Kumar KP, Singh A. Multicentric Trial of Diabecon (D-400)-A Herbomineral Preparation on Lipid Profile in Diabetes Mellitus. *International Journal of Diabetes in Developing Countries*. 1996;16:87-9.
- [107] Boral G, Bandopadhyaya G, Boral A, Das N, Nandi P. Geriforte in anxiety neurosis. *Indian Journal of Psychiatry*. 1989;31(3):258.
- [108] Sahu M, Bhat R, Kulkarni K. Clinical evaluation of himplasia in benign prostatic hyperplasia: an open clinical trial. *Med Update*. 2003;11:75-8.

- [109] Tewiri P, Kulkiro MKS. A study of lukol in leucorrhoea, pelvic inflammatory diseases and dysfunctional uterine bleeding. *Ancient Science of Life*. 2001;21(2):139.
- [110] Sahu M, Gupta S, Srivastava P. Effect of Renalka syrup in urinary tract infection. *Indian Pract*. 2002;55:101-6.
- [111] Rehman A, Adnan M. Nutritional potential of Pakistani medicinal plants and their contribution to human health in times of climate change and food insecurity. *Pakistan Journal of Botany*. 2018;50(1):287-300.
- [112] Mahmood N, Muazzam MA, Ahmad M, Hussain S, Javed W. Phytochemistry of *Allium cepa* L.(Onion): An overview of its nutritional and pharmacological importance. *Scientific Inquiry and Review*. 2021;5(3):41-59.
- [113] Hussain S, Tanvir M, Ahmad M, Munawar KS. Phytochemical Composition of Mint (*Mentha*), its Nutritional and Pharmacological Potential. *Lahore Garrison University Journal of Life Sciences*. 2021;5(04):241-58.
- [114] Singh N, Jha A, Chaudhary A, Upadhyay A. Enhancement of the functionality of bread by incorporation of Shatavari (*Asparagus racemosus*). *Journal of food science and technology*. 2014;51(9):2038-45.
- [115] Mehta M. Development of low cost nutritive biscuits with Ayurvedic formulation. *International Journal of Ayurvedic and Herbal Medicine*. 2013;3(3):1183.
- [116] Singh AK, Srivastava A, Kumar V, Singh K. Phytochemicals, medicinal and food applications of Shatavari (*Asparagus racemosus*): An updated review. *The Natural Products Journal*. 2018;8(1):32-44.
- [117] Kumar S, Mehla R, Dang A. Use of shatavari (*Asparagus racemosus*) as a galactopoietic and therapeutic herb—a review. *Agricultural Review*. 2008;29(2):132-8.
- [118] Patel A, Kanitkar U. *Asparagus racemosus* willd--form bordi, as a galactagogue, in buffaloes. *The Indian veterinary journal*. 1969;46(8):718-21.
- [119] Dhuria R, Sharma T, Karnani M, Rai K, Singh S. Effect of herbs as feed additives on nutrient utilization in rams. *Indian Journal of Animal Nutrition*. 2013;30(3):281-4.
- [120] Kumar S, Mehla R, Gupta A, Meena R. Influence of *Asparagus racemosus* (Shatavari) supplementation during different stage of lactation on estrus behavior and reproductive performance in Karan Fries crossbred cows. *Livestock Research for Rural Development*. 2010;22(5):99.

Edytor – Michał Nowicki

Received: 23.02.2023

Accepted: 25.03.2023

Shabbir Hussain

Institute of Chemistry

Khwaja Fareed University of Engineering and Information Technology

Rahim Yar Khan 64200, Pakistan

e-mail: shabchem786@gmail.com

Mob # +92-3214140130